



## Blank qPCR Master Mix (2x)

### Kit Components

Component	Cat. No. E0403-01 100 reactions of 25 µl	Cat. No. E0403-02 200 reactions of 25 µl	Cat. No. E0403-03 1000 reactions of 25 µl
Blank qPCR Master Mix (2x)	1 x 1.25 ml	2 x 1.25 ml	10 x 1.25 ml
UNG (uracil-N-glycosylase) 1 U/µl	30 µl	55 µl	270 µl
25 mM MgCl <sub>2</sub>	0.2 ml	0.4 ml	2 ml
Water, nuclease free	1 x 1.25 ml	2 x 1.25 ml	10 x 1.25 ml

## Blank qPCR Master Mix (2x), plus ROX Solution

### Kit Components

Component	Cat. No. E0404-01 100 reactions of 25 µl	Cat. No. E0404-02 200 reactions of 25 µl	Cat. No. E0404-03 1000 reactions of 25 µl
Blank qPCR Master Mix (2x)	1 x 1.25 ml	2 x 1.25 ml	10 x 1.25 ml
ROX Solution, 25 µM	55 µl	110 µl	530 µl
UNG (uracil-N-glycosylase) 1 U/µl	30 µl	55 µl	270 µl
25 mM MgCl <sub>2</sub>	0.2 ml	0.4 ml	2 ml
Water, nuclease free	1 x 1.25 ml	2 x 1.25 ml	10 x 1.25 ml

### Storage

Store at -20°C for long-term storage or at 4°C for up to 1 month.

This product is developed, designed and sold exclusively for research purposes and in vitro use only.

EURx Ltd. 80-297 Gdańsk Poland ul. Przyrodników 3, NIP 957-07-05-191, KRS 0000202039  
www.eurx.com.pl, orders@eurx.com.pl, tel. +48 58 524 06 97, fax +48 58 341 74 23

## Description

- Blank qPCR Master Mix (2x) is a universal solution for quantitative real-time PCR and two-step real-time RT-PCR and can be used on most real-time PCR cyclers available.
- The master mix does not contain any dye and allows the user to choose a dye and optimize the dye concentration.
- The master mix contains Perpetual Taq DNA Polymerase, optimized reaction buffer and dNTPs (dTTP is partially replaced with dUTP).
- Perpetual Taq DNA Polymerase is recombinant Taq DNA Polymerase bound to anti-Taq monoclonal antibodies that block polymerase activity at moderate temperatures.
- The polymerase activity is restored during the initial denaturation step when amplification reactions are heated at 95°C for at least two minutes.
- Use of the “hot start” enzyme prevents extension of misprimed products and primer-dimers during reaction setup leading to higher specificity and sensitivity of PCR reactions.
- The polymerase enables convenient room temperature reaction setup.
- Blank qPCR Master Mix (2x) contains dUTP, which partially replaces dTTP. It allows the optional use of a uracil-N-glycosylase (UNG) to prevent carryover contamination between reactions. UNG removes uracil from any dU-containing contaminating amplicons, leaving abasic sites and making DNA molecules susceptible to hydrolysis during the initial denaturation step.
- There are two variants of the kit: without ROX and with ROX Solution provided separately. The use of ROX passive reference dye is necessary for all real-time PCR cyclers from Applied Biosystems and optional for cyclers from Stratagene. ROX compensates for variations of fluorescent signal between wells due to slight differences in reaction volume and fluorescence fluctuations. ROX is not involved in PCR reaction, has a different emission spectrum than SYBR Green I and does not interfere with real-time PCR on any instrument. Refer to the table below to determine the recommended amount of ROX (25 µM) required for a specific PCR cycler.

## Recommended amounts of ROX for a specific real-time PCR cycler

Instrument	Amount of ROX per 25 µl reaction	Amount of ROX per 1.25 ml of 2x master mix	Final ROX concentration
Applied Biosystems: 7300, 7900HT, StepOne, StepOnePlus, ABI PRISM	0.5 µl	50 µl	500 nM
Applied Biosystems: 7500, ViiA 7, Stratagene: Mx3000P, Mx3005P,	0.5 µl 10 x diluted (in water)	50 µl 10 x diluted (in water)	50 nM
PCR machines from other manufacturers: Bio-Rad, Roche, Corbett, Eppendorf, Cepheid, etc.	Not required	Not required	-

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## Protocol

### Preparation of PCR Reaction:

Component	Volume/reaction	Final concentration
Blank qPCR Master Mix (2x)	12.5 $\mu$ l	1 x 2.5 mM MgCl <sub>2</sub>
Forward Primer	Variable	0.3-0.5 $\mu$ M
Reverse Primer	Variable	0.3-0.5 $\mu$ M
Template DNA	Variable	$\leq$ 500 ng
qPCR dye	Variable	Variable
Optional: ROX Solution, 25 $\mu$ M	0.5 $\mu$ l or 0.5 $\mu$ l 10 x diluted	500 nM 50 nM
Optional: UNG (uracil-N-glycosylase) 1 U/ $\mu$ l	0.25 $\mu$ l	0.25 U/reaction
Optional: 25 mM MgCl <sub>2</sub>	0-1.5 $\mu$ l	2.5-4 mM
Water, nuclease free	To 25 $\mu$ l	-
Total volume	25 $\mu$ l	-

### Notes:

1. A reaction volume of 25  $\mu$ l should be used with most real-time cyclers. Other reaction volumes may be used if recommended for a specific instrument.
2. Optimal amplicon length in real-time PCR is 70-200 bp.
3. Thaw, gently vortex and briefly centrifuge all solutions.
4. Set up PCR reactions at room temperature. Use of Blank qPCR Master Mix (2x) allows room temperature reaction setup.
5. Prepare a reaction master mix by adding all the reaction components except template DNA.
6. Mix the reaction mix thoroughly and dispense appropriate volumes into PCR tubes or plates.
7. Add template DNA/cDNA ( $\leq$ 500 ng/reaction) to the individual PCR tubes or wells containing the reaction mix. For two-step RT-PCR, the volume of cDNA added should not exceed 10% of the final PCR volume.
8. Centrifuge briefly to settle down the reaction components and remove bubbles. Bubbles interfere with fluorescent detection.
9. Place the samples in the cycler and start the program.
10. Standard concentration of MgCl<sub>2</sub> in real-time PCR reaction is 2.5 mM (as provided with the 1 x Blank qPCR Master Mix). In most cases this concentration will produce optimal results. However, if a higher MgCl<sub>2</sub> concentration is required, use 25 mM MgCl<sub>2</sub> stock solution (provided) and add to a reaction.
11. A final primer concentration of 0.3-0.5  $\mu$ M is usually optimal, but can be individually optimized in range of 0.1  $\mu$ M to 1  $\mu$ M. The recommended starting concentration is 0.5  $\mu$ M. Raising primer concentration may increase PCR efficiency, but negatively affect PCR specificity. Optimal primer concentration depends on the individual reaction and the real-time PCR cycler used.
12. Readjust the threshold value for analysis of every run.
13. If using Bio-Rad iCycler iQ or MyiQ instruments collect well factors at the beginning of each experiment. Use an external well factor plate according to the manufacturer's recommendations. Well factors are used to compensate for any excitation or pipetting variations.

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## Thermal Cycling Conditions:

Step	Temperature	Time	Number of Cycles
Optional: UNG pre-treatment	50°C	2 min	1
Initial Denaturation	95°C	10 min	1
Denaturation	94°C	15 s	35-45
Annealing	50-60°C	30 s	
Extension	72°C	30 s	
Optional: Data acquisition	X°C	15 s	
Cooling	4°C	Indefinite	1

### Notes:

1. The incubation step of 50°C for 2 minutes must be added if a uracil-N-glycosylase is used to prevent carryover contamination. UNG degrades any dUMP-containing PCR products.
2. During the initial denaturation step UNG and antibodies that block Taq DNA Polymerase are inactivated. The anti-Taq antibodies and UNG require at least 2 min or 10 min incubation at 95°C, respectively. When UNG is not used in PCR reaction the duration of the initial denaturation step can be reduced to 2-5 min at 95°C.
3. UNG activity may be partially restored at temperatures lower than 55°C due to refolding. It is recommended to perform PCR using a temperature equal 55°C or above for the annealing step. After completing the PCR cool reactions to 4°C and load directly on a gel or store frozen.
4. Melting curve analysis should be performed to verify the specificity and identity of PCR products. Melting curve analysis is an analysis step built into the software of real-time cyclers. Melting curve data between 65°C and 95°C should be acquired.
5. Data acquisition should be performed during the extension step. To suppress fluorescence readings caused by the generation of primer-dimers an additional data acquisition step can be added to the protocol. It is possible when  $T_m$  of primer-dimers differs from  $T_m$  of the specific product ( $T_m$  are generated during melting curve analysis). The temperature of the data acquisition step should be above  $T_m$  of primer-dimers but approximately 3°C below the  $T_m$  of the specific product.
6. Always check the PCR product specificity by gel electrophoresis when designing a new assay. Melting temperatures of the specific product and primer-dimers may overlap,

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