

## NheI

5'-GCTAGC-3'  
3'-CGATCG-5'

Cat. No.	size
E2294-01	500 units
E2294-02	2500 units

**Reaction Temperature:** 37°C

**Inactivation Temperature (20 min):** 65°C

**Prototype:** NheI

**Source:** *Neisseria mucosa heidelbergensis*

**Note 1:** Recombinant. Purified from an *E.coli* strain carrying the NheI gene from *Neisseria mucosa heidelbergensis*.

**Package Contents:**

- NheI
- 10 x ONE Buffer
- BSA [100x]  
Added as separate component to prevent reaction buffer precipitation.
- Dilution Buffer # NheI  
Added for enzymes exceeding 10 U/μl in concentration. High protein concentration warrants optimal stability during prolonged storage. Use dilution buffer to prepare short term working stocks (5-10 U/μl, non-freezing at -20°C).

**Storage Conditions:** Store at -20°C.

**Double Digestion – Buffer Compatibility:**

ONE Buffer is compatible with most EURx restriction enzymes.

**Restriction Enzyme Buffer Compatibility:**

Both, enzyme and buffers are fully compatible to restrictases and buffer systems from other manufacturers and can be used along in double digestions. To obtain best results, consult the corresponding manuals of all involved products.

**DNA Methylation:**

No Inhibition: dam, dcm, EcoKI

Potential Inhibition: CpG

### Standard Reaction Protocol (for 50 μl volume):

**Mix** the following reaction components:

- 1-2 μg pure DNA or 10 μl PCR product (≈0.1-2 μg DNA)
- 5 μl 10 x ONE Buffer
- 0.5 μl BSA [100x]
- 1-2 U NheI (use 1 U per μg DNA, < 10% React. Volume!)

*Tips:* Add enzyme as last component. Mix components well before adding enzyme. After enzyme addition, mix gently by pipetting. Do not vortex. High (excess) amounts of enzyme can greatly speed up the reaction.

add sterile H<sub>2</sub>O to 50 μl final volume

**Incubate** for 1 h at 37°C

To obtain complete digestion of high molecular weight DNA, (e.g. plant genomic DNA), add excess amounts of enzyme and prolong the incubation time.

**Stop** reaction by alternatively

- Addition of 2.1 μl EDTA pH 8.0 [0.5 M], final 20 mM or
- Heat Inactivation  
20 min at 65°C or
- Spin Column DNA Purification  
(e.g. EURx PCR/DNA Clean-Up Kit, Cat.No. E3520) or
- Gel Electrophoresis and Single Band Excision  
(e.g. EURx Agarose-Out DNA Kit, Cat.No. E3540) or
- Phenol-Chloroform Extraction or Ethanol Precipitation.

### Non-optimal buffer conditions:

To compensate for the lack of enzyme activity, increase the amount of enzyme and/ or reaction time accordingly. The following values may serve as orientation:

- Enzyme amount:** Instead of 1 U enzyme, use ~4 U of enzyme in buffers providing 25% rel. activity, ~2 U in 50%, ~1.5 U in 75% or ~1 U in 100%, respectively.
- Reaction time:** Increase by ~1.3-fold (75% rel. activity), ~2-fold (50%) or ~4-fold (25%).

### Unit Definition:

One unit is the amount of enzyme required to completely digest 1 μg of Lambda DNA (HindIII digest) in 1 hr. Total reaction volume is 50 μl. Enzyme activity was determined in the recommended reaction buffer.

**Note 2:** To avoid star activity it is not recommended:

- to use more than 100 units per reaction;
- to incubate over 4 hours.

**Note 3:** Salt concentration above 50 mM inhibits NheI activity.

### Reaction Buffer:

1 x ONE Buffer

To be supplemented with 100 μg/ml bovine serum albumin.

### Storage Buffer:

10 mM Tris-HCl (pH 7.5 at 25°C), 1 mM dithiothreitol, 250 mM NaCl, 0.1 mM EDTA, 0.1% (v/v) Tergitol™ TMN, 500 μg/ml bovine serum albumin and 50% (v/v) glycerol.

### Quality Control:

All preparations are assayed for contaminating endonuclease, 3'-exonuclease, and nonspecific single- and double-stranded DNase activities.

This product is developed, designed and sold exclusively for research purposes and in vitro use only.

EURx Ltd. 80-297 Gdańsk Poland ul. Przyrodników 3, NIP 957-07-05-191, KRS 0000202039  
www.eurx.com.pl, orders@eurx.com.pl, tel. +48 58 524 06 97, fax +48 58 341 74 23